



Complete Genome Sequences of *Mandrillus leucophaeus* and *Papio ursinus* Cytomegaloviruses

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The complete genome sequences of *Mandrillus leucophaeus* and *Papio ursinus* cytomegaloviruses were determined. An isolate from a drill monkey, OCOM6-2, and an isolate from a chacma baboon, OCOM4-52, were subjected to pyrosequencing and assembled. Comparative alignment of published primate cytomegaloviruses (CMVs) showed variable sequence conservation between species.

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ytomegaloviruses (CMVs) are members of the Betaherpesvirinae subfamily and infect most primates, including humans, Old World monkeys (OWM), and New World monkeys (NWM) (1). CMVs are considered to be host restrictive due to the lack of evidence of cross-species infection and are thought to have coevolved with their host. Nonhuman-primate CMVs are the closest relatives to human CMV (HCMV) (2). Drill monkeys and baboons are infected in the wild and are also used as models for CMV vaccine and antiviral development (2). To solidify the evolutionary relationship between primate CMVs and HCMVs, a drill monkey CMV (DrCMV) isolate, OCOM6-2 (3), and a chacma baboon CMV (BaCMV) isolate, OCOM4-52 (4), were sequenced using a Roche GS Junior (454 Life Sciences, a Roche company, Branford, CT). The long read lengths (median, 380 nucleotides [nt]) allowed the complete CMV genomes to be de novo assembled using the 454 Assembly Suite, version 2.9, without manual intervention. CLC Genomics Workbench version 7.0.4 (CLC Bio-Qiagen, Aarhus, Denmark) was used for validation. A total of 82,066 reads were used for the DrCMV isolate with a Q40PlusBases score of 99.01%. A total of 111,885 reads were used for the BaCMV isolate with a Q40PlusBases percentage of 98.78% of all nucleotides. Annotations were transferred from the cercopithecine herpesvirus 5 strain 2715 (CHV5) (NCBI reference sequence NC_012783), using the Rapid Annotations Transfer Tool (RATT) (5) and verified using CLC Genomics Workbench version 7.0.4. CHV5 is a species-specific CMV most closely related to the sequenced isolates. Using the multiple sequence alignment program MAFFT shows that DrCMV and BaCMV share 78%, human CMV and chimpanzee CMV share 60%, and BaCMV and rhesus monkey CMV share 58% sequence identity across their entire genomes (6).

Nucleotide sequence accession numbers. The annotated sequences for *Mandrillus leucophaeus* isolate OCOM6-2 and *Papio ursinus* isolate OCOM4-52 cytomegaloviruses have been deposited in GenBank under the accession numbers KR297253 and KR351281, respectively.

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REFERENCES

- 1. Davison AJ, Holton M, Dolan A, Dargan DJ, Gatherer D, Hayward GS. 2013. Comparative genomics of primate cytomegaloviruses, p 1–22. *In* Reddehase M (ed), Cytomegaloviruses: from molecular pathogenesis to intervention, vol 1. Caister Academic Press/Horizon, Norfolk, United Kingdom.
- Fruh K, Malouli D, Oxford KL, Barry PA. 2013. Non-human-primate models of cytomegalovirus infection, prevention, and therapy, p 463–496. *In* Reddehase M (ed), Cytomegaloviruses: from molecular pathogenesis to intervention, vol 2. Caister Academic Press/Horizon, Norfolk, United Kingdom.
- Blewett EL, Lewis J, Gadsby EL, Neubauer SR, Eberle R. 2003. Isolation of cytomegalovirus and foamy virus from the drill monkey (*Mandrillus leucophaeus*) and prevalence of antibodies to these viruses amongst wildborn and captive-bred individuals. Arch Virol 148:423–433. http:// dx.doi.org/10.1007/s00705-002-0937-9.
- Blewett EL, White G, Saliki JT, Eberle R. 2001. Isolation and characterization of an endogenous cytomegalovirus (BaCMV) from baboons. Arch Virol 146:1723–1738. http://dx.doi.org/10.1007/s007050170059.
- Otto TD, Dillon GP, Degrave WS, Berriman M. 2011. RATT: Rapid Annotation Transfer Tool. Nucl Acids Res 8:e57 http://dx.doi.org/10.1093/ nar/gkq1268.
- Katoh K, Misawa K, Kuma K-I, Miyata T. 2002. MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. Nucleic Acids Res 30:3059–3066. http://dx.doi.org/10.1093/nar/gkf436.