RESEARCH ARTICLE

Tumor Markers and Signatures

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A prospective cohort study identifies two types of HIV+ Kaposi Sarcoma lesions: proliferative and inflammatory

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Abstract

Kaposi sarcoma (KS) is the most common cancer in people living with HIV (PLWH) in many countries where KS-associated herpesvirus is endemic. Treatment has changed little in 20 years, but the disease presentation has. This prospective cohort study enrolled 122 human immunodeficiency virus (HIV) positive KS patients between 2017 and 2019 in Malawi. Participants were treated with bleomycin, vincristine and combination antiretroviral therapy, the local standard of care. One-year overall survival was 61%, and progression-free survival was 58%. The 48-week complete response rate was 35%. RNAseq (n = 78) differentiated two types of KS lesions, those with marked endothelial characteristics and those enriched in inflammatory transcripts. This suggests that different KS lesions are in different disease states consistent with the known heterogeneous clinical response to treatment. In contrast to earlier cohorts, the plasma HIV viral load of KS patients in our study was highly variable. A total of 25% of participants had no detectable HIV; all had detectable KSHV viral load. Our study affirms that many KS cases today develop in PLWH with well-controlled HIV infection and that different KS lesions have differing molecular compositions. Further studies are needed to develop predictive biomarkers for this disease.

KEYWORDS

clinical study, differential gene expression, Kaposi sarcoma herpesvirus, transcriptome RNA sequencing

Abbreviations: BV, bleomycin/vincristine; cART, combination antiretroviral therapy; CEMIP, cell migration inducing hyaluronidase 1; CR, complete response; DTG/3TC/TDF, dolutegravir/ lamivudine/tenofovir; ECFC, endothelial colony-forming cells; EFV/3TC/TDF, efavirenz/lamivudine/tenofovir; HIV, human immunodeficiency virus; IL-1 β , interleukin-1 β ; KCH, Kamuzu Central Hospital; KICS, KSHV Inflammatory Cytokine Syndrome; KS, Kaposi sarcoma; KSHV, Kaposi sarcoma-associated herpesvirus; KS-IRIS, KS-Immune Reconstitution Inflammatory Syndrome; LMIC, low- and middle-income countries; LTFU, loss-to-follow up; MCD, Multicentric Castleman Disease; OS, overall survival; PAM, partitioning around medoids; PD, progressive disease; PEL, primary effusion lymphoma; PFS, progression-free survival; PLWH, people living with HIV; PR, partial response; RPKM, reads per kilobase of exon per million reads mapped; SD, stable disease; TIVE, telomerase-immortalized microvascular endothelial cells; tSNE, t-distributed stochastic neighbor embedding.

Razia Moorad and Edwards Kasonkanji contributed equally to this study.

What's new?

Kaposi sarcoma (KS) is a common cancer among people living with HIV, particularly in countries where KS-associated herpesvirus (KSHV) is endemic. In this study, the authors analyzed skin biopsies and identified transcript patterns in HIV-positive KS patients in Malawi, Africa, to better characterize the clinical and molecular features of KS. In total, 78 transcriptomes were generated from patient biopsies, and two clusters of KS lesions were detected. Many patients had undetectable HIV viral load, though all had measurable levels of KSHV. The findings indicate that HIV-positive individuals develop KS lesions with distinct molecular characteristics, potentially enabling the identification of predictive biomarkers.

1 | INTRODUCTION

Kaposi sarcoma (KS) is among the most common cancers in Eastern and Central Africa.^{1,2} In Malawi, in 2021, KS accounted for approximately one-fifth of all cancer cases.³ Incidence rates are similar in Uganda and other KS-endemic countries.^{4,5} In South Africa, KS is the most common cancer in men living with HIV.⁶ In KS-endemic countries, KS affects human immunodeficiency virus (HIV)-positive and negative individuals, males and females and children.

First described in 1872. KS preceded the introduction of HIV into the human population. In the US and Europe, one-third of KS cases develop in PLWH on stable combination antiretroviral therapy (cART), who have suppressed HIV viral loads and near normal CD4 counts.^{7,8} Classic KS was described at a time when a detailed assessment of human immune status was not possible: it clustered in specific populations, such as men of Mediterranean descent.⁹ Classic KS today is not necessarily associated with overt immune deficiency either.¹⁰ In KSendemic countries, high CD4 counts provide less protection against the development of KS in persons living with HIV (PLWH) compared to non-KS-endemic countries,^{11,12} an indication that HIV-KS will continue to be of concern in Sub-Saharan Africa (SSA). Potential reasons for the high incidence rates of KS in the region include the high rates of Kaposi Sarcoma-associated herpesvirus (KSHV), KSHV/HIV coinfection, suboptimal cART coverage or co-infection of KSHV with other diseases.¹²

KSHV infection necessarily precedes KS disease. KSHV is also the etiological agent for diseases that often co-exist in KS patients, such as Multicentric Castleman Disease (MCD), Primary Effusion Lymphoma (PEL), KS-Immune Reconstitution Inflammatory Syndrome (KS-IRIS) and KSHV Inflammatory Cytokine Syndrome (KICS; reviewed in Ref. 13). In SSA, KSHV seroprevalence exceeds 80% before puberty. Here, KSHV infection often precedes HIV acquisition (except in instances of HIV mother-to-child transmission). Other childhood infections, such as malaria, are common and may modulate KSHV acquisition.¹⁴

Many HIV-positive KS patients with limited-stage disease respond to combination antiretroviral therapy (cART) alone initially; however, most KS patients require concurrent or subsequent cytotoxic cancer therapy,¹⁵⁻¹⁷ including those not infected with HIV (classic KS). Standard treatment strategies for KS are based on cytotoxic chemotherapy.¹⁸⁻²² Prior studies^{16,23-27} informed first-line treatment recommendations in SSA and other low- and middle-income countries (LMIC), including vincristine/bleomycin (BV) and non-liposomal doxorubicin or paclitaxel. HIV-KS is treated with pegylated liposomal doxorubicin in high-income countries.²⁸⁻³¹ Overall survival (OS) and progression-free survival (PFS) in SSA are lower than in the United States or Europe.²⁷ This discrepancy is due to a combination of factors, including late presentation and insufficient access to pegylated liposomal doxorubicin (Doxil). It is unknown whether, in addition, there also exist differences in the underlying biology of KS, such as KSHV strain distribution³² or if there are different molecular drivers of the disease. This represents a gap in our knowledge and a barrier to optimal treatment designs that our study aimed to address.

We report on a prospective cohort study of KS patients treated at the Kamuzu Central Hospital (KCH) in Lilongwe, Malawi. The motivation was that a better understanding of both clinical and molecular parameters would inform disease management and uncover novel targets for intervention. The aim was to describe KS in PLWH with access to cART and medical care typical for countries with endemic KS, that is, childhood-acquired KSHV³³ and epidemic HIV. The primary objectives were to define the baseline characteristics, OS, PFS, and complete response (CR) at 48 weeks. All participants received bleomycin (15 IU/ m²) and vincristine 2 mg (fixed dose) and initiated cART concurrently or were continued on cART. The number of chemotherapy cycles was left to the clinician's discretion per the local standard of care. RNAsequencing of 78 KS skin lesion biopsies was performed to identify KS subsets based on human transcript patterns. This study produced the largest database of KS transcriptome data to date.

2 | METHODS

2.1 | Study design

LCCC1424 was a prospective, open-label cohort study of pathologyconfirmed HIV-associated KS patients that initiated chemotherapy treatment in Malawi. Patient eligibility was defined as having histological confirmation of KS, HIV positive, on or off cART, 18 years of age or older, residing within 200 km from the study site, and having the ability to give informed consent. Pregnancy and breastfeeding were not considered exclusion criteria, given that the study was observational and with diagnostic and treatment interventions administered according to local standards of care. Prior cART experience was allowed, but previous chemotherapy was not. Participants with previously treated KS were excluded. Eligible participants were enrolled from February 2017 to June 2019 and were followed for up to 96 weeks. Upon histological confirmation of KS diagnosis, a comprehensive baseline evaluation was performed. The baseline evaluation included the collection of two skin biopsies and baseline clinical laboratory testing. KS staging was assessed by a complete skin examination and chest radiograph, conforming to previously published criteria.³⁴ Five marker lesions were selected and evaluated at each visit. Marker lesions were selected based on the operating procedures from concurring randomized phase III KS studies at this site.^{16,27} Specifically, we identified lesions that were at least 1 cm \times 1 cm, at least 1 cm away from, and distinguishable from any nearby lesions. We then choose the five largest lesions with clearly distinguishable margins. These were saved on a body map to ensure the same lesions were measured each time.

The decision to initiate chemotherapy treatment was at the physician's discretion. All but one participant started treatment at screening. One participant was delayed in initiating treatment as the participant did not promptly report back to the clinic. Here, the enrollment date was used as the initiation date. Chemotherapy was administered according to a standardized clinic treatment algorithm and consisted of BV only, while cART was administered according to national guidelines. Treatment toxicity was graded using the National Cancer Institute Common Technology Criteria for Adverse Events (CTCAE) version 5.

2.2 | Statistical analysis

The Kaplan-Meier method was used to estimate the OS and PFS, with the two-sided 95% confidence intervals (95% Cls) calculated using the log-log transformation. A Cox proportional-hazards regression model was used to estimate the hazard ratios for multiple clinical variables. Multivariate logistic regression was used to study associations between the dichotomous CR outcome and targeted clinical variables at 48 weeks. The associations between KSHV viral load and the targeted clinical variables were performed independently. The Pearson or the Spearman methods were used to calculate the correlations for continuous clinical variables. The binary clinical variables were studied using either the Wilcoxon rank sum test (minimum group size <30) or the *t*-test (minimum group size \geq 30). The normality assumption was verified by the Shapiro-Wilk test.

2.3 | KSHV viral load

DNA was extracted using the MagNA Pure Instrument (Roche). Following DNA extraction, KSHV viral load was determined as described.³⁵

2.4 | Immunohistochemistry

Formalin-fixed paraffin-embedded blocks containing the skin biopsies were sectioned at the UNC Pathology core facility. Sections were stained for LANA following a previously described protocol³⁶

3

(Figure S1). A dilution factor of 1:100 of mouse anti-human LANA antibodies was used (Leica Biosystems Cat# NCL-L-HHV8-LNA).

2.5 | Transcriptome analysis

Approximately 30 mg tissue was processed using the Qiagen, RNeasy Fibrous Tissue Mini Kit (Qiagen, cat# 74704), with 300 µL RLT, 0.05% DX Reagent, and one 3.0 mm stainless steel ball. The sample was homogenized for 3 minutes or until completion using the Qiagen Tissue Lyser (Qiagen, cat# 85300), and RNA was isolated according to the manufacturer's instructions. RNA quantity and quality were verified by Qubit assay (Life Technologies) and on a 4200 Tape Station (Agilent Technologies). Library preparation and templating were automated on the Ion Chef System (Life Technologies Pub. No. MAN0013432 Rev. H.0), where libraries were diluted to 75 pM before templating on the Ion 540 chip (Thermofisher cat#. A27765). Sequencing was performed on the Ion Torrent S5 sequencer (Life Technologies Pub. No. MAN0010811 Rev: C.0). The cleaned FASTQ files are submitted to SRA achieves. The sequencing coverage and quality statistics for each sample are summarized in Table S1.

Human transcription was determined using CLC Genomics Workbench version 21.0.5 (Qiagen) and further processed using DESeq2.³⁷ Protein coding genes with at least 10 sequence reads in at least one sample and expressed in at least half of the participants were retained for analysis. Sequence reads were randomly subsampled to a maximum of two million reads for the cell line data (SRP035883 and SRP078245). Normalized reads were further transformed using the Variance Stabilizing Transformations (VST) implemented in DESeq2. The 2000 most variable genes (based on their coefficient of variation) were used for hierarchical clustering based on the Euclidean distance matrix and Ward's linkage using R, version 4.1.2.

3 | RESULTS

3.1 | Baseline characteristics

Prospective participants were screened between February 2017 and June 2019. Fifteen participants were excluded due to non-KS histology. One participant was excluded due to being HIV-negative at enrollment, and another one due to having a previous treatment of KS less than a year before enrollment. Fifteen participants were withdrawn due to incomplete enrollment. Two participants withdrew after one treatment cycle and another after four treatment cycles due to suspected side effects. The study had 122 participants with complete clinical data (Table 1).

The median age was 36 (IQR: 32-44) years; 98 (80%) participants were male. A total of 18 (15%) participants were classified as tumor severity TO, and 104 (85%) as T1; 67 (55%) participants had an illness severity score of S1, and 55 (45%) were classified as S0. A total of

TABLE 1 Participant baseline characteristics at enrolment for a total number of participants (n = 122; interquartile range [IQR]; standard deviation [SD]).

Total number of participants (n)	122
Age, years, median (IQR)	36 (32-44)
Gender; men, n (%)	98 (80)
Staging	
Tumor severity TO (ACTG), n (%)	18 (15)
Karnofsky performance status ≤70%, n (%)	43 (35)
Illness severity S1, n (%)	67 (55)
Symptoms	
Edema present, n (%)	60 (51)
Visceral disease present, n (%)	8 (7)
Oral involvement present, n (%)	41 (35)
HIV Characteristics	
Knew HIV+ diagnosis before KS diagnosis, n (%)	64 (53)
Months HIV+ diagnosis known, median (IQR) (n = 61)	16 (6-57)
On ART prior to KS diagnosis, n (%)	61 (50)
Months on ART prior to diagnosis, median (IQR) (n = 61)	16 (6-57)
CD4 count, median (IQR) (n $=$ 115)	197 (96-337)
HIV viral load log copies/mL/median (IQR) $(n = 116)$	3 (2-5)
HIV viral load suppressed <1000 copies/mL, n (%) (n = 116)	67 (58)
Baseline Lab Results	
Hemoglobin, g/dL/mean (SD)	11 (3)
Platelets, 10 ³ /µL/median (IQR)	234 (157-313)
White blood cell, $10^3/\mu$ L/median (IQR)	5 (4-6)
Absolute neutrophil count, 10 ³ /μL/median (IQR)	2 (1-3)
Creatinine, mg/dL/median (IQR)	0.8 (0.7-0.9)
Bilirubin, mg/d/median (IQR) (n = 111)	0.4 (0.3-0.5)

43 (35%) participants had a Karnofsky performance status (KPS) ≤70. Presenting symptoms included edema in 60 (49%), visceral disease in 8 (7%), and oral involvement in 41 (34%) participants. Before diagnosis, 64 (52%) participants were aware of being HIV infected for a median of 15.7 months (IQR: 6-57.1). A total of 61 (50%) participants were on cART before their KS diagnosis for a median of 15.7 months (IQR: 6-57.1). The median CD4 count was 197 (IQR: 96-337) cells/µL, and the median HIV viral load was 2.6 log10 copies/mL (IQR: 1.6-4.7). A total of 67 (58%) participants had an HIV viral load of <1000 copies/mL. For participants who were on cART before screening, the CD4 count was 212 (IQR: 98.5-345.2, n = 56) cells/µL; for those not on cART, it was 176 (IQR: 75.5-332, n = 59) cells/µL. For participants who were on cART before screening, the HIV genome copy number was 1.60 (IQR: 1-3.3, n = 57) log10 copies/mL, and for those not on cART, it was 3.39 (IQR: 2.1-4.9, n = 59) log10 copies/mL. **TABLE 2** Clinical outcomes for the total number of participants (n), interquartile range (IQR), and Bleomycin/Vincristine (B/V).

Variable	Sample size (n)	Value
Treatment cycles	122	
Cycles received, median (IQR)		16 (6-17)
Received ≥16 cycles (%)		66 (54)
Reason treatment not completed (%)	52	
Died		23 (44)
Defaulted		24 (46)
Moved to another HIV treatment facility		2 (4)
Toxicity		2 (4)
Disease Progression		1 (2)
Treatment stock out (%)	122	
Had no missed doses due to stock out (B or V)		62 (51)
Had 1 missed dose due to stock out		13 (11)
Had >1 missed dose due to stock out		47 (39)
Drugs missed due to stock out (%)	122	
Had 1 missed dose of bleomycin		14 (12)
Had >1 missed dose of bleomycin		22 (18)
Had 1 missed dose of vincristine		8 (7)
Had >1 missed dose of vincristine		28 (23)
Adverse events during treatment (%)	122	
Grade 2 anemia		14 (12)
Grade 3/4 anemia		4 (3)
Grade 2 neutropenia		19 (16)
Grade 3/4 neutropenia		12 (10)
Grade 2 thrombocytopenia		1 (0.8)
Grade 3/4 thrombocytopenia		0 (0)
Non-hematologic grade 3/4 event		0 (0)
Had at least one delayed or reduced dose due to adverse event		0 (0)
Vital status (%)	122	
Alive		67 (55)
Died		33 (27)
Lost to follow-up		22 (18)
48 weeks response (%)	99	
Complete response		35 (35)
Partial response		28 (28)
Stable disease		4 (4)
Progressive disease		32 (32)

3.2 | Outcomes

The treatment outcomes are summarized in Table 2. The median number of treatment cycles was 16 (IQR: 7-17), with 66/122 (54%) participants receiving 16 or more treatment cycles. A total of 52/122 (43%) participants discontinued treatment early, receiving

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less than 16 treatment cycles without achieving a complete response (CR); 23 (44%) died, and one (2%) experienced progressive disease (PD) after six treatment cycles. Twenty-four (46%) of the 52 participants defaulted after a median of six (IQR: 2-9) treatment cycles, two participants were transferred to new facilities, and two dropped out due to suspected treatment toxicity. Sixty (49%) participants had experienced at least one delayed or missed dose of either chemotherapy agent due to stock out. The median number of cycles for participants with one missed or delayed dose was 3 (IQR: 2-4). Fourteen (12%) participants had at least one cycle without bleomycin (median missed doses 2 [IQR: 1-3]), and 22 (18%) participants had at least one cycle without vincristine (median missed doses 3 [IQR 2-4]). This was an observational study with treatment determined by the treating physicians according to hospital guidelines (often dictated by local drug availability and patient needs). Most patients received chemotherapy every 2 weeks, which could be delayed for adverse events such as anemia, neutropenia, thrombocytopenia, or other grade 3 to 4 adverse events. The minimum period between cycles was never less than 14 days.

The adverse effects include grade two anemia in 19 (16%) participants and grade 3/4 anemia in 4 (3%) participants. Grade 2 neutropenia occurred in 19 (16%) participants, and grade 3/4 neutropenia occurred in 12 (10%) participants. One participant had grade four bleomycin-induced dermatitis. There were no detected cases of grade 3 or 4 lung toxicity, neuropathy or thrombocytopenia.

Outcome definitions were harmonized with response category definitions (complete response, partial response [PR], stable disease, progressive disease) based on two concurring randomized phase III ACTG/AMC KS studies at this site.^{16,27} The definitions below were specific to assessing the five KS marker lesions. CR was the absence of any detectable residual disease, including tumor-associated edema, persisting for at least 4 weeks. In patients known to have had visceral disease, an attempt at restaging with appropriate radiographic procedures should be made. PR was defined as no new oral lesions or new or progressive visceral involvement, the appearance or worsening of tumor-associated edema or effusions or the development of five or more new cutaneous marker lesions in anatomic sites which were previously documented as having no evidence of cutaneous disease; and a 50% decrease in the sum of the products of the largest perpendicular diameters of the five cutaneous marker lesions compared to entry. PD was defined as follows: for participants with <50 cutaneous marker lesions 25% increase in the sum of the products of perpendicular diameters of the five cutaneous marker lesions compared to entry. For participants with ≥50 cutaneous marker lesions, 25% increase in the sum of the products of the perpendicular diameters of the five cutaneous marker lesions compared to entry.

The censoring date for the assessment of the vital statuses of the participants was 30 June 2020. At censoring, 67 (55%) participants were alive, 33 (27%) participants were dead and 22 (18%) participants were loss-to-follow-up (LTFU). Of the 33 deaths in this set, 12 were due to PD, 12 were treatment-related and 4 were due to other causes unrelated to treatment or KS. The cause of death could not be ascertained for five participants.

Among the participants with an evaluable response (n = 99), 35 (35%) achieved a CR at 48 weeks based on clinical assessment and

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28 (28%) achieved a PR. Eight (8%) participants achieved stable disease (SD); 28 (28%) experienced PD. For the 69 patients who were not evaluated at exactly 48 weeks (missed appointment), the response was imputed from either the last visit that preceded the 48-week follow-up or the first visit after the 48-week follow-up, whichever was closer. Participants with baseline KPS <70 were less likely to achieve CR at 48 weeks ($P \le .01$, n = 99). No other parameters (age, gender, tumor, illness severity, edema, visceral disease, oral involvement, knew they were HIV+ before KS diagnosis, number of months HIV+ diagnosis was known, CD4 count, HIV viral load, hemoglobin, platelets, white blood cells, creatinine or bilirubin) were significantly associated with response (Figure S2). All participants received cART concurrent to chemotherapy according to national standards and local availability. Twenty (16%) participants received dolutegravir/lamivudine/tenofovir (DTG/3TC/TDF). A total of 91 (74%) participants received efavirenz/lamivudine/tenofovir (EFV/3TC/TDF). Eleven participants received other cART regimens. OS and PFS were estimated using the Kaplan-Meier method (Figure 1A,B). At 1 year, the OS was 61% (95% CI: 69%-85%), and PFS was 58% (95% CI: 63%-80%), with LTFU considered a censoring event. Four variables had a statistically significant association with OS. The first variable was sex (HR = 5.19, 95% CI: 1.0-27): males were five times more likely to die than females: however, four times more males than females were enrolled in the study. The second variable was oral involvement (HR = 0.19, 95%) CI: 0.04-0.96). The third variable was CD4 count (HR = 0.89, 95%CI: 0.80-0.99). The fourth variable was increased hemoglobin (HR = 0.79, 95% Cl: 0.65-0.97). For each unit of hemoglobin increase (g/dL), the hazard of death decreased by 21%. No statistically significant (based on $P \leq .05$ after multivariate adjustments) relationships were identified between PFS hazard and any of the clinical variables. For patients with no prior cART at baseline, no statistically significant associations were made with survival outcomes due to the small sample size and, thus, lack of statistical power. Overall, almost half of the participants died within one year of diagnosis.

3.3 | KSHV and HIV viral loads

KSHV and HIV genome copy numbers (cps/mL) were determined for 111 participants at baseline (Figure 1C). We compared the HIV and KSHV genome copy numbers obtained in this enrollment period (2017-2019) to our prior study of KS patients in Malawi that were enrolled between 2008 to 2010.³⁸ In the cART naive 2008 to 2010 population, KSHV viral load (copies/mL, log10 scale) was 3.1 ± 0.84 (mean \pm SD, n = 111). The KSHV viral load was similar in cART and chemo-naive KS patients in 2017 to 2019 at 3.2 ± 0.82 (mean \pm SD, n = 69). KSHV viral load was independent of CD4 count (r^2 = .003 by the linear model, $P \le .61$ by *F*-test) in both time frames. HIV genome copy number at baseline was trimodal in the current study: 40% of KS participants had no detectable HIV in their blood (limit of detection at 50 cps/mL), 35% had between 50 and 50 000 cps/mL and 25% of KS participants presented with >50 000 cps/mL. Our study thus reports a significant change to the monomodal distribution of HIV genome



FIGURE 1 Survival curves were estimated by the Kaplan-Meier method (A) OS (n = 122) and (B) PFS (n = 122). There was a total of 27 events for OS and 33 events for PFS. (C) A comparison of HIV and KS viral loads and CD4 counts from a previous study (2008-2010) and our study (2017-2019). (D) Boxplot of KSHV Viral load (copies/mL, log10 scale) of subgroups defined by awareness of HIV status at least 3 months before KS diagnosis (variable name = "Knew HIV status," n = 111; $P \le .04$). [Color figure can be viewed at wileyonlinelibrary.com]

copies, which was 5.0 ± 0.60 (mean \pm SD) log10 cps/mL in the KS patients from 2008 to 2010. There was a trend for lower CD4 counts associated with higher HIV genome copy number ($r^2 = .18$ by the linear model, $P \le 10^{-5}$ by *F*-test); however, many KS patients had ≥ 400 CD4 cells/µL. In sum, a quarter of KS seen in Malawi today is no longer associated with uncontrolled HIV viral load or CD4 depletion.

Each clinical variable was tested for association with the KSHV genome copy number at baseline in a multivariate analysis (Figure 1D). First, participants who were not aware of their HIV status at least 3 months before their KS diagnosis had significantly higher KSHV viral loads (3.40 ± 0.83 ; mean \pm SD, n = 52) compared to participants who were aware of their HIV status (3.08 ± 0.78 ; mean \pm SD, n = 59, $P \le .05$ by *T*-test). Second, higher hemoglobin levels (g/dL) were associated with lower KSHV viral loads, but the strength of the relationship was small (Spearman correlation coefficient r = -.2, $P \le .002$). Our study did not identify significant associations between CR, HIV viral loads (copies/mL, log10 scale), KSHV viral loads (copies/mL, log10 scale) and CD4 counts for participants who were HIV-positive/cART naive for more than 3 months or those who had edema at baseline.

3.4 | Human transcriptome sequencing identified two subtypes of KS

Targeted RNAseq yielded 95 human transcriptomes from KS biopsies with matched clinical data. Sixteen samples with low sequencing depth were removed from subsequent analyses to avoid threshold sensitivity biases, resulting in 78 samples with comparable technical quality and total cellularity. The clinical characteristics of this set were not different from the overall cohort. A total of 11 467 genes expressed in at least half the samples with a mean expression >10 reads per kilobase of exon per million reads mapped (RPKM) were retained for analysis.

To determine how different the transcriptome of KS biopsies was from "normal" endothelial cells, we incorporated data from DiMaio et al.,³⁹ who described circulating endothelial colony-forming cells (ECFC) of lymphatic and blood origin, and data from Sychev et al.,⁴⁰ who determined the transcriptome in Telomerase-immortalized Microvascular Endothelial Cells (TIVE) cells in both KSHV infected and uninfected cells. The t-distributed stochastic neighbor embedding (tSNE) (Figure 2A-C) projection of samples colored by the unsupervised hierarchical clustering based on the 2000 most differentially regulated genes easily distinguish the uninfected and KSHV-infected pure endothelial cells (colored gray and red, respectively) from the KS biopsies (colored yellow or blue). The transcripts most differentially regulated between the KS lesions and the "normal" endothelial cells include, for instance, matrix metalloproteinase MMP1, an established marker of angiogenesis. Significantly downregulated genes in the KS lesions, as compared to cell lines, were DCBLD2, RELN, FTH1, BCAR1, APLN, APOLD1 and LSS. The upregulated genes in this set of KS lesions, as compared to cell lines, included S100A12, a calciumbinding protein linked to inflammation and cancer and adiponectin (ADIPOQ), an adipokine that regulates glucose levels and the breakdown of fatty acids. Other significantly upregulated genes included



FIGURE 2 (A-C) tSNE visualizations of the 2 KS patient clusters (yellow and blue), typical uninfected endothelial cell lines (gray), and KSHV-infected TIVE cell lines (red). Both visualizations display distinct separation of normal and KSHV-infected endothelial cells vs KS patient clusters. [Color figure can be viewed at wileyonlinelibrary.com]

GPD1, HBB, FCGR3B, SLC4A1, ALAS2 and HBD. In the absence of single-cell data, we cannot decide whether the difference in gene transcription represents differences in the KSHV-positive tumor cells to KSHV-negative and positive endothelial cells or is the result of mixed cellularity of the patient biopsies, as compared to pure cell lines.

Next, unsupervised hierarchical clustering was performed on the KS lesion data alone. Four potential KS tumor subtypes were identified, but due to the small sample size, we focused the analyses on the two most highly divergent subgroups of KS (Figure 3A). The first subtype included 35 samples (blue), and the second subtype 43 samples (yellow). To investigate the molecular differences between these two subtypes of KS lesions, genes were clustered using the partitioning around medoids (PAM) method. Figure 3B shows a volcano plot of adjusted significance in relation to the relative mean difference in gene expression between the two clusters, blue and yellow, using the blue cluster as the reference group. The volcano plot highlights individual differentially expressed genes of >8-fold mean difference between the two clusters. These were interleukin-1 β (IL-1 β) and



FIGURE 3 (A) A heatmap of the top 2000 most variable genes (median-centered VST transformed data from DESeq2) between the two KS subtypes. (B) Volcano plot depicting the differentially expressed genes within the two KS subtypes (blue cluster as the reference group). [Color figure can be viewed at wileyonlinelibrary.com]

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CXCL5, indicative of an inflammatory microenvironment in the yellow subtype, and KIAA1199/CEMIP (cell migration inducing hyaluronidase 1). CEMIP regulates hyaluronic acid, an extracellular matrix component, and has been previously associated with KSHV infection.⁴¹ Other significantly upregulated transcripts include VAT1L, LY6H, RBP4 and PPP1R1A. ANXA8L1, Annexin A8 Like 1 protein, was the only downregulated gene in the yellow cluster and, by implication, upregulated in the blue cluster. We provisionally term the yellow cluster the "inflammatory KS subtype" and the blue cluster the "proliferative KS subtype." Note that unsupervised clustering is driven by the sum total of all changes in gene transcription, even if any one gene alone does not rise to individual significance in this particular experiment. No significant differences were seen between the two tumor subtypes in complete response, OS or PFS. In sum, transcriptional profiling demonstrates substantial heterogeneity among KS lesions, with at least two major subtypes, one characterized by an inflammatory profile and the other reflecting predominantly endothelial cell components.

4 | DISCUSSION

The risk of KS is increased substantially in PLWH. In the US, which has low KSHV prevalence in the general population, the risk for KS in PLWH is 100-fold above that of the general population despite 20 years of cART.^{6,42} BY comparison, the standardized incidence ratio of developing cervical cancer, considered AIDS-associated in the original CDC definition, for PLWH is in the 2-fold range, driven essentially by the large, near-universally high HPV prevalence.⁴³ In Sub-Saharan Africa, where KSHV general population prevalence is above 50%, the estimated relative risk of developing KS in PLWH is numerically lower; however, there are many more PLWH and many more people infected with KSHV. In SSA, KS is among the five most prevalent cancers overall, irrespective of HIV status and irrespective of sex. We believe this is due to KSHV seroprevalence rates in the region being as high as 70%,⁴⁴ while in the United States and Europe, KSHV seroprevalence is lower and concentrated in PLWH.⁴⁵

The US standard of care, using single-agent liposomal doxorubicin (Doxil), was developed 20 years ago.^{15,28,46} Doxil is neither consistently available nor affordable in many public sector hospitals in SSA. Rather BV and, more recently, paclitaxel, in combination with cART, are used.^{12,24,27} Whether pomalidomide, which was fast-tracked based on a 60% overall response rate in 28 treatment-experienced KS patients,⁴⁷ will become widely affordable in SSA remains to be seen. Studies with other "imids"⁴⁸ and a second study with pomalidomide⁴⁹ support the clinical efficacy of this agent class in KS.

Here, we presented data from an observational, prospective cohort study of PLWH who present with KS aimed to define the clinical characteristics of KS patients and their responses to BV plus cART, the Malawi national standard of care. The goal was to be as inclusive as possible and to characterize a prototypical, contemporary population of PLWH who develop KS. This contrasts with prior interventional trials, which may screen out as many as half of KS patients. A total of 122 biopsy-confirmed KS cases were enrolled between February 2017 and June 2019. All participants had to be KS treatment-naive; approximately half were on cART before enrollment (median of 15 months). This distribution sets the current study apart from earlier studies.^{16,27} It represents the typical population of KS patients in recent times, that is, after the 2016 adoption of WHO guidelines for immediate cART treatment of all HIV+ persons irrespective of CD4 count or clinical symptoms. The OS was 61% at 1 year, and the PFS was 58%. At 48 weeks, CR was achieved in 35% and PR in 28% (n = 99). The median number of treatment cycles received was 16; however, half of the participants received suboptimal chemotherapy due to drug shortages, which remains common in LMIC environments.

The OS and PFS were in line with prior studies. Bower et al.¹⁵ reported a 5-year OS of 83% using cART/liposomal Doxorubicin (Doxil) in a 140-person cohort representing a 1989 to 2013 UK population of 89% cART naive KS patients. This is among the highest survival times reported to date in KS. None of the data from SSA approaches this result. A randomized phase III trial comparing the efficacy of BV plus cART to paclitaxel plus cART in advanced KS patients (stage T1) without prior cART obtained a 1-year PFS of 44% (N = 132) and 1-year OS of 80% in the BV plus cART group.²⁷ A 2003 to 2009 trial by Mosam et al.²³ reported a 1-year OS of 74% (n = 53) using BV/cART plus doxorubicin in T1 stage KS. Crudely approximating across studies and collection times, OS and PFS for KS in SSA have not changed in the last 20 years though admittedly, the data is minimal and often confounded by loss to follow-up.

All patients had detectable KSHV in the blood. Significantly elevated KSHV viral loads were observed in participants who did not know their HIV status at least 3 months before the study, reaffirming that HIV-induced immune suppression and/or HIV directly reactivates KSHV.

Prior studies consistently observed the existence of two HIV-KS lesion subtypes based on the KSHV gene transcription.^{38,50} Limited, that is, latent KSHV transcription was the predominant subtype; cART seemed to restrict viral transcription to the latent genes and viral miRNAs compared to KS lesions from cART-naive patients. This is consistent with in situ analyses, which always detect the LANA protein, but rarely other viral proteins in KS lesions.⁵¹ Here, we queried human gene transcription in KS lesions assembling the most extensive set of KS transcript data today. Differential gene expression identified two subtypes of KS lesions; one enriched in genes that induce the proliferation and migration of cells and a second enriched in inflammatory genes. Because of the targeted design for human genes only, we could not relate viral to human gene transcription.

We did not observe a difference in the survival curves or any statistically significant association with response hazard between the two KS tumor subtypes. This was expected for multiple reasons. Even though our study reports on more samples than prior studies, ⁵²⁻⁵⁵ this remains a small data set with limited outcome data. It represents a snapshot of KS for a specific time and place. Second, despite multiple KS lesions on the same patient, we could biopsy only one lesion at baseline due to IRB concerns. This under-sampling likely obscured any

dominant transcription pattern required to predict survival outcomes. The situation in KS contrasts with most solid tumors, where the primary tumor can be discerned and biopsied and where the primary tumor subtype predicts response. For KS, by contrast, we know that different lesions on the same patient represent different overt classes (patch, plaque, nodular), have different histological compositions,⁵⁶ and respond differently to systemic treatment.

We tried to identify potential associations between clinical characteristics and gene expression within one single lesion (out of many possible) on a patient's skin. We did not find any. This may be due to the limited power of our study. Larger studies in terms of participants and length of follow-up may uncover those. Alternatively, this may be due to the biology of KS, where single skin lesions develop independently, and it is their aggregation, as well as the lesions that develop internally in the lung and liver, as well as co-occurring MCD and PEL that drive the most robust clinical characteristics.^{57,58} Unfortunately, institutional review boards frown upon extended experimental biopsies. We estimate that one would need at least as many biopsies as marker lesions to establish robust linkages between overall systemic clinical characteristics and tumor gene expression.

There are limitations to our study. Ascertaining CR for KS tumors accurately requires biopsies of all indicator lesions. That was not feasible in this and other LMIC settings. To date, most IRBs will no longer approve studies requiring extensive biopsies. We were worried that insisting on this "gold" standard may introduce an enrollment bias, as many potential participants are averse to multiple biopsies. Our study aimed to capture everyone with KS symptoms. Thus, the CR numbers reported here may be overly optimistic from a strictly scientific point of view. The participants and their providers reported an improvement. Given the smoldering, remitting/recurring nature of KS in the presence of cART, further studies, including quality of live measures, are indicated to determine the most meaningful response criteria in this patient population today.

The principal limitation in calculating survival outcomes was the lack of events and LTFU after 1 year, which reduced the statistical power in detecting late differences in survival outcomes. Out of the 78 participants with matched high-quality human transcriptome data, only eight deaths and 27 PD could be ascertained. This was expected partly due to the liberal inclusion criteria of including "all comers" and partly due to the limited resources at the site. In addition, our study had a short follow-up. It was a single-center experience compared to two recent multicenter, randomized phase III clinical trials that reported lower survival for HIV-associated KS in the SSA.^{16,27} Considering that our study aimed to explore survival outcomes under the local standards of care, there were limitations in drug supply, treatment interruptions and patient retention. Under these circumstances, and including the molecular heterogeneity of KS lesions, we submit that PFS and OS are more robust clinical measures than clinical response measurements developed for US AIDS patients before the availability of cART.³⁴

In sum, the standard of care for HIV-associated KS in the region where KS and HIV burden is the highest globally lags behind the best possible care available in the United States and Europe. Our study underscores the molecular diversity of individual KS lesions and KS patients. Yet, we still lack robust biomarkers to predict individual KS risk for patients on cART or individual therapy responses.

AUTHOR CONTRIBUTIONS

The work reported in the article has been performed by the authors unless clearly specified in the text. Razia Moorad: Investigation, Data Curation, Formal Analysis, Writing - Original Draft. Edwards Kasonkanji: Data Curation, Formal Analysis. Joe Gumulira: Investigation. Yolanda Gondwe: Project Administration. Morgan Dewey: Investigation. Yue Pan: Data Curation, Formal Analysis. Alice Peng: Data Curation, Formal Analysis. Linda J. Pluta: Investigation. Evaristar Kudowa: Data Curation. Richard Nyasosela: Investigation. Tamiwe Tomoka: Investigation. Hannock Tweya: Investigation. Tom Heller: Investigation. Salem Gugsa: Investigation. Sam Phiri: Conceptualization, Project Administration. Dominic T. Moore: Formal Analysis. Blossom Damania: Conceptualization, Funding Acquisition, Writing - Review & Editing. Matthew Painschab: Writing - Review & Editing, Supervision. Mina C. Hosseinipour: Conceptualization, Writing - Review & Editing, Project Administration, Supervision. Dirk P. Dittmer: Conceptualization, Methodology, Formal Analysis, Project Administration, Funding Acquisition, Validation, Supervision, Writing - Review & Editing.

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CONFLICT OF INTEREST STATEMENT

Dirk P. Dittmer has been a consultant to Thermo Fisher Scientific. The other authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

The data supporting our study's findings are openly available in the NCBI BioProject Database (https://www.ncbi.nlm.nih.gov/bioproject), linked to the BioProject accession number PRJNA947563. Other data that support the findings of our study are available from the corresponding author upon request.

ETHICS STATEMENT

The study was approved by the Malawi National Health Science and Research Committee (NHSRC) and the Institutional Review Board (IRB) at the University of North Carolina at Chapel Hill. A separate informed consent for tissue collection was obtained in specific languages modeled after previous and existing cancer studies.

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10

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SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

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