

## MMRRC UNC – Genotyping Protocol

<b>MMRRC Strain ID</b>	17240
<b>MMRRC Strain Name</b>	CB17.Cg-Igh-J <sup>tm1(3H9-VDJ)Mwg</sup> Prkdc <sup>scid</sup> /Mmnc
<b>Gene Name(s)</b>	immunoglobulin heavy chain, joining region (Igh-J) protein kinase, DNA activated, catalytic polypeptide (Prkdc)
<b>Breeding Protocol(s)</b>	Sib-mating
<b>Protocol Date</b>	9/7/13

### MMRRC #17240 PCR Reactions

Genotyping for this strain includes 3 reactions: one four-primer reaction for scid, and two two-primer reactions for H1 and 3H9/56R. The 101 bp of WT band cannot be shown in the four-primer reaction. A two-primer reaction using the primers MB 575 + MB 576 is needed to confirm the homozygous samples.

	<u>1X</u>
ddH <sub>2</sub> O	13
5X Buffer	5.0
25mM MgCl <sub>2</sub>	2
10mM dNTPs	0.5
10 μM Primer F	1
10 μM Primer R	1
Taq	0.5
DNA	2

#### **Thermal Cycler:**

Step 1: 94°C for 5 min  
 Step 2: 94°C for 30 sec  
 Step 3: 61 °C for 30 sec  
 Step 4: 72°C for 45 sec  
 Step 5: 35x from step 2 to step 4  
 Step 6: 72°C for 7 min

#### **Taq: Apex and Chromataq 5X Buffer**

**Primer sequences 5' to 3':** Primers are 10 μM with respect to each primer.

MB 575: GAGAAAAGGAGGATCATG GATTCAAGAAATAAATGTAACG

MB 576: TGGCCCCTGCTAACTTTCTCTTAGCA

MB 577: TGGTATCCACAACATAAAATACGCTAA

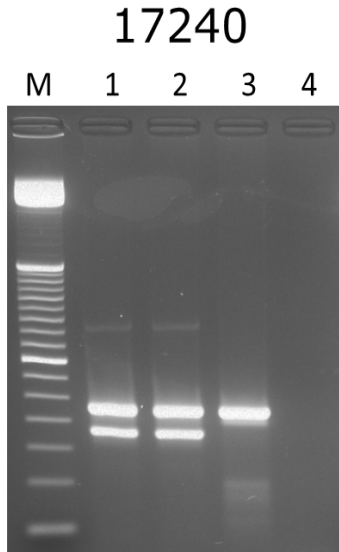
M-MB 578: CCTAAGAGTCACTTTCTCCATCTACACAGTGAAGTGCC

**Bands expected:** MB 575 + MB 576 WT: 101bp

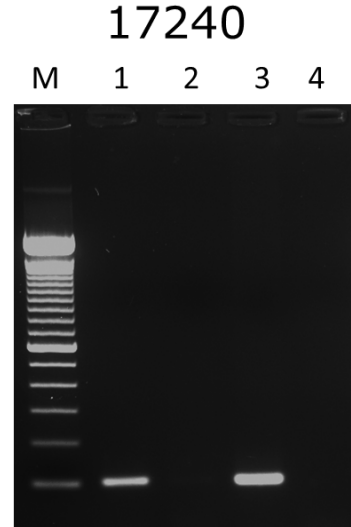
MB 577 + M-MB 578 scid band: 180 bp

MB 575 + M-MB 578 Common band: 229 bp

Run on 3% agarose gel in TAE.



Primers: MB 575 + MB 576 + MB 577 + M-MB 578  
 Lane 1 and 2: Hom; Lane 3: WT; Lane 4: H<sub>2</sub>O  
 M: 50 bp DNA ladder Invitrogen



Primers: MB 575 + MB 576  
 Lane 1 and 3: WT; Lane 2: H<sub>2</sub>O; Lane 4: Hom;  
 M: 100 bp DNA ladder Invitrogen

### MMRRC #17240 PCR – JH1 Reaction

	<u>1X</u>
ddH <sub>2</sub> O	13
5X Buffer	5.0
25mM MgCl <sub>2</sub>	2
10mM dNTPs	0.5
10 μM Primer F	1
10 μM Primer R	1
Taq	0.5
DNA	2

#### **Thermal Cycler:**

Step 1: 94°C for 5 min  
 Step 2: 94°C for 30 sec  
 Step 3: 59 °C for 30 sec  
 Step 4: 72°C for 1 min  
 Step 5: 34x from step 2 to step 4  
 Step 6: 72°C for 7 min

**Taq: Apex and Chromataq 5X Buffer**

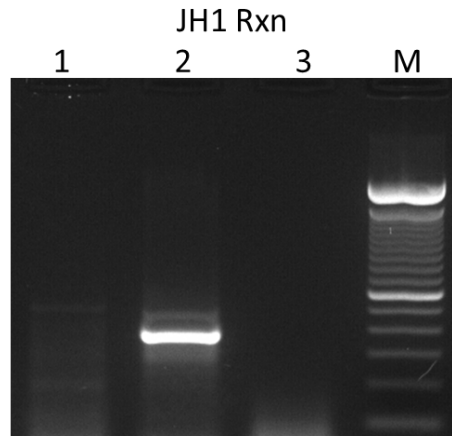
**Primer sequences 5' to 3':** Primers are 10 μM with respect to each primer.

MB 631: GCC AAG GAC TTA CCA AGA GG

MB 632: 5GAT GCA GGA CTC ACC TGA CC

**Bands expected:** WT: 376 bp  
 Tg+: No product

Run on 1% agarose gel in TAE.



Lane 1: Tg+; Lane 2: WT; Lane 3: H<sub>2</sub>O; M: 100 bp DNA ladder (Invitrogen)

### MMRRC #17240 PCR – 3H9 / 56R Reaction

	<u>1X</u>
ddH <sub>2</sub> O	13
5X Buffer	5.0
25mM MgCl <sub>2</sub>	2
10mM dNTPs	0.5
10 μM Primer F	1
10 μM Primer R	1
Taq	0.5
DNA	2

**Thermal Cycler:**

- Step 1: 94°C for 5 min
- Step 2: 94°C for 30 sec
- Step 3: 50°C for 30 sec
- Step 4: 72°C for 45 sec
- Step 5: 34x from step 2 to step 4
- Step 6: 72°C for 7 min

**Taq: Apex and Chromataq 5X Buffer**

**Primer sequences 5' to 3'**: Primers are 10  $\mu$ M with respect to each primer.

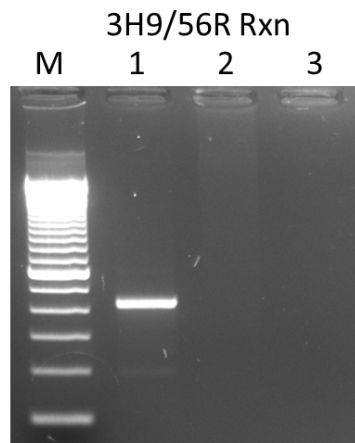
MB 722: CTG TCA GGA ACT GCA GGT AAG G

MB 730: AGT CCA TAA CAT AGG AAT ATT TAC TC

**Bands expected:** Tg+: 400 bp

WT: No band

Run on 1% agarose gel in TAE.



Lane 1: Tg+; Lane 2: WT; Lane 3: H<sub>2</sub>O; M: 100 bp DNA ladder (Invitrogen)