**Alkaline lysis mini preps:**

1. grow 3ml culture O/N in LB
2. Spin 5 minutes at 4000g, or remove 1.5ml and spin at top speed for 1 min.
3. Aspirate almost all of LB
4. “Washboard” style break up pellet
5. Add 200ul P1 to fully R/S
6. Add 200ul P2, invert 6 times and wait 1 minute, or swirl repeatedly for 1-2 minutes
7. Add 280ul P3, invert 10 times or or swirl repeatedly for 1-2 minutes
8. Decant into 1.5ml eppie
9. Spin top speed at 4C for 10 minutes
10. Remove sup with barrier tip pipette and place into clean eppie
11. Add 680ul of isopropanol
12. Vortex
13. Spin at top speed in table top cetrifuge at RT for 10 minutes
14. Aspirate isopropanol mix
15. Add 1ml of 70% EtOH
16. Aspirate all of remaining EtOH
17. R/S in 30ul of diH20 (sequencing compatible)

**Buffer P1** - Resuspension Buffer

50mM Tris-Cl, pH 8.0, 10mM EDTA, 100ug/mL RNase A

Storage condition - 4oC after adding RNase A

Prep - Dissolve 6.06g Tris base, 3.72g EDTA-2H20 in 800mL dH20. Adjust the pH to 8.0 with HCl. Adjust the volume to 1 liter with dH2O. Add 100mg RNase A per liter of P1.

**Buffer P2** - Lysis Buffer

200mM NaOH, 1% SDS

Storage condition - RT

Dissolve 8.09g of NaOH pellets in 950mL dH2O, 50mL 20% SDS solution. The final volume should be 1 liter.

**Buffer P3** - Neutralization Buffer for Qiatips, Midiprep, Maxiprep, and Gigaprep kits.

3.0M potassium acetate, pH 5.5

Storage condition - 4C or RT

Dissolve 294.5g potassium acetate in 500mL dH2O. Adjust the pH to 5.5 with glacial acetic acid (about 110mL). Adjust the volume to 1 liter with dH2O.